Genos: A Human-Centric Genomic Foundation Model

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Abstract

- 2 The rapid expansion of human genomic data demands foundation models that manage ultra-
- 3 long sequences and capture population diversity, limitations common in existing models
- 4 which lack human-specific representation and clinical inference efficiency. Here, we
- 5 introduce Genos (Genos-1.2B/Genos-10B), a human-centric genomic foundation model
- 6 engineered for million-base-pair sequence modeling. Genos utilizes a large-scale Mixture of
- 7 Experts (MoE) structure, optimized for a 1Mb context, trained on high-quality human de
- 8 novo assemblies from datasets such as HPRC and HGSVC, representing diverse global
- 9 populations. A suite of optimization strategies was implemented to ensure training stability
- and enhance computational efficiency, which collectively reduces costs and facilitates
- 11 million-base-pair context modeling. Functionally, Genos performs single-nucleotide
- resolution analysis and dynamically simulates the cascade effects of non-coding variations on
- 13 RNA expression profiles. In comprehensive evaluations, Genos uniformly surpasses State-of-
- 14 the-Art models on critical human genomics benchmarks and demonstrates robust Omics-Text
- 15 cross-modal diagnostic capabilities. We present a systematic technical evaluation and
- validation of Genos's architecture, training convergence, and performance across standard
- benchmarks. This work provides a reliable technical blueprint and performance benchmark
- 18 for the development of the next generation of high-efficiency genomic foundation models.
- 19 Genos model weights, inference code, and usage documentation are publicly available on
- 20 GitHub (https://github.com/BGI-HangzhouAI/Genos) and Hugging Face Hub
- 21 (https://huggingface.co/BGI-HangzhouAI), with additional cloud services accessible via BGI
- 22 DCS Cloud—all released under the MIT License.

1. Introduction

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1.1 The Paradigm Shift: Genomics and Foundation Models

- 26 Genomics research is currently transitioning from an early phase of massive data accumulation to the
- 27 contemporary era of intelligent analysis and insight extraction. The proliferation of high-throughput
- 28 sequencing technologies has generated an unprecedented volume of nucleic acid sequence data,
- 29 making deep learning-based Genomic Foundation Models (GFMs) a crucial computational tool for
- deciphering the complexity of life. Analogous to Large Language Models (LLMs) in Natural
- 31 Language Processing, GFMs aim to learn the intrinsic "grammar" and "semantics" of the genome
- through large-scale pre-training, enabling unified analysis of functional element identification, variant
- pathogenicity prediction, and phenotype regulatory networks. This technological breakthrough is
- 34 pivotal for accelerating precision medicine and population health research.

1.2 Significance of Genos in the Field

- 36 Significant progress has been made in the GFM landscape, with seminal works like EVO2 [1] and
- 37 AlphaGenome [2] leading the trend toward long-sequence modeling and cross-species generalization.
- 38 However, when these models are applied to human translational medicine and clinical high-
- 39 throughput analysis, they encounter two core bottlenecks.
- 40 Bottleneck I: The Human-Centric Representational Gap. The OpenGenome2 dataset used by EVO2
- 41 prioritizes cross-species coverage over population diversity, leading to systematic bias in the
- representation of human-specific regulatory elements (e.g., enhancers, promoters) and rare variants.
- 43 Similarly, AlphaGenome relies on cohorts with limited reference genomes, struggling to accurately
- 44 capture complex population-specific genetic patterns. This fundamentally restricts the models'
- 45 predictive accuracy and generalizability in complex human disease and rare disorder research.
- 46 Bottleneck II: Efficiency and Deployment Challenges for Ultra-Long Sequences. While existing
- 47 models have achieved context modeling up to the million-base-pair (1Mb) scale, this often incurs
- 48 prohibitive computational costs. For instance, the 40B-parameter version of EVO2 requires extensive
- 49 GPU clusters for training and exhibits high inference latency, unsuitable for time-sensitive clinical
- analysis. Furthermore, specialized architectures often lack modularity, making them incompatible
- with mainstream cloud computing infrastructures, significantly raising the barrier to deployment and
- 52 broad application.
- Considering the two core bottlenecks, we focused our efforts on robust data engineering and an
- 54 optimized technical architecture. On the data side, we curated a human-centric, multi-source dataset,
- integrating high-quality, haplotype-resolved assemblies from the Human Pangenome Reference
- 56 Consortium (HPRC) [3-5] and the Human Genome Structural Variation Consortium (HGSVC) [6] to
- 57 ensure robust cross-ethnic generalizability. Architecturally, we rooted our design in an evolved
- Transformer framework, augmenting it with a Mixture-of-Experts (MoE) [7] structure to address the
- 59 computational challenges of modeling sequences up to a million bases. This was achieved by
- integrating elements such as Rotary Position Embedding (RoPE) [8] for extreme context lengths, and
- 61 multiple parallelism strategy (including Tensor, Pipeline, Context, Data, and Expert Parallelism) to
- 62 ensure stable and efficient large-scale training. This comprehensive work on data and architecture,
- 63 coupled with extensive training and optimization, culminated in the release of Genos (Genos-
- 64 1.2B/Genos-10B), a human-centric Genomic Foundation Model.

Genos stands at the forefront of genomic foundation models, playing a pivotal role in the field of genomics. It has the potential to revolutionize multiple aspects of genomic research and its applications. In precision medicine, Genos can analyze an individual's genomic data to predict disease risks with greater accuracy. For instance, by identifying key genetic markers associated with diseases such as cancer or neurodegenerative disorders, facilitates the development of personalized treatment regimens. This not only improves the effectiveness of treatment but also reduces the risk of adverse

71 reactions to medications.

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- In the realm of group health monitoring, by analyzing genomic data from large populations, the model
- 74 facilitates the precise identification genetic trends within different ethnic groups, which is crucial for
- understanding the genetic basis of diseases prevalent in specific populations. These critical genomic
- 76 insights provide the scientific foundation necessary for formulating can be used to develop targeted
- 77 preventive measures and healthcare policies. In developmental biology, Genos can help in
- values of the genetic mechanisms underlying embryo development. By analyzing the genomic
- sequences at different stages of development, researchers can uncover how genes are regulated to
- 80 drive the formation of various tissues and organs.

1.3 Objectives and Core Design Feature of Genos

- 82 The objective for Genos is to provide a genomic intelligence analysis engine characterized by superior
- accuracy and efficiency, thereby advancing the field into a mass application phase. Genos provides
- 84 significant methodological advancements.
- 85 In data processing, Genos integrates standardized, high-quality data from leading international
- 86 genomics initiatives, including the Human Pangenome Reference Consortium (HPRC) [3-5] and the
- Human Genome Structural Variation Consortium (HGSVC) [6]. By constructing a multi-source,
- 88 heterogeneous genomic dataset spanning global populations and incorporating hundreds of nearly
- 89 telomere-to-telomere (T2T) assemblies, Genos achieves robust cross-ethnic generalizability. To ensure
- 90 the reliability and representativeness of training data, we designed a multi-stage quality control
- 91 pipeline that progressively filters out intergenic sequences of varying lengths, many of which contain
- 92 segmental duplication (SD) regions.
- 93 The model's architecture is rooted in an evolved Transformer [9] framework, augmented by a
- 94 Mixture-of-Experts (MoE) [7] structure. This design effectively overcomes the long-standing
- 95 computational challenge associated with modeling sequences that exceed a million bases. The
- 96 integration of ultra-long sequence parameterization, multi-dimensional parallel computing, and
- 97 specialized complementary attention mechanisms allows Genos to perform single-nucleotide
- 98 resolution modeling on ultra-long sequences. Consequently, this provides a more comprehensive
- analytical depth, allowing for the precise capture and analysis of fine-scale genetic details across the
- 100 entire genome.
- 101 Functionally. Genos has the core ability to accurately identify key functional elements in the genome.
- 102 It can deeply analyze the cascade effect of micro-gene variation on the transcriptional regulatory
- 103 network. This is a significant improvement over traditional methods, which often have limitations in
- 104 predicting regulatory elements in the non-coding region. Genos is capable of single-nucleotide
- 105 resolution analysis within ultra-long non-coding regions and can dynamically simulate the cascade

effect of variation sites on RNA expression profiles, offering a novel paradigm for molecular

mechanism analysis.

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2. Methodology

2.1 Data Collection and Preprocessing

- 110 The training data for Genos were curated from multiple high-quality genomic sources, including 231
- 111 haplotype-resolved assemblies from the HPRC (release 2), 65 assemblies from the HGSVC, and 21
- genomes from the Centre d'Etude du Polymorphisme Humain (CEPH) cohort, along with two
- reference genomes, GRCh38 and CHM13. In total, the dataset comprises 636 high-quality genomes,
- 114 representing diverse global populations. Each genome sequence was processed using a one-hot
- tokenizer, with a vocabulary consisting of the four canonical nucleotides (A, T, C, G), the
- undetermined base N, and special tokens such as <EOD> marking sequence boundaries. No cell-type
- 117 specific labels, epigenetic features (e.g., histone modifications), or other functional annotations were
- incorporated during this stage. This ensures that the model learns a general-purpose representation of
- the human genome, unbiased towards any particular biological context or experimental condition.
- 120 Training was performed in two major stages. In the pre-training stage, samples from HPRC release 2
- were divided into four groups at an approximate 3:3:3:1 ratio, corresponding to sequence lengths of
- 122 8,192 bp, 32,768 bp, 131,072 bp, and 1,024,000 bp. Within each stage, about one-quarter of the
- samples had both haplotypes reverse-complemented, while the remaining samples retained the
- forward strand orientation. Samples from HGSVC and CEPH pedigrees were all processed into 8,192
- bp fragments, with one-quarter of them reverse-complemented in the same manner. Both reference
- genomes (GRCh38 and CHM13) were prepared with both forward and reverse strands at every length
- 127 scale. To reduce non-informative intergenic content, 8,192 bp fragments excluded regions located
- more than 5,120 bp away from any gene boundary, while 32,768 bp fragments excluded regions
- beyond 10,240 bp from gene boundaries. The four pre-training datasets were then sequentially
- introduced to the model by increasing sequence length, resulting in a total of approximately 1.4
- trillion (1,400B) tokens. In the subsequent continued pre-training (CPT) stage, the same samples were
- reshuffled across lengths and strand orientations to generate an additional 2.6 trillion (2,600B) tokens,
- which were further randomized before being fed into the model.
- 134 Crucially, it is important to emphasize that all filtering was discontinued in the subsequent Continued
- 135 Pre-Training (CPT) stage. The additional 2.6 trillion tokens used in CPT were generated from the
- original samples without any intergenic distance-based exclusion. This ensured that the model was
- 137 extensively exposed to and trained on distal intergenic regions, segmental duplications, transposable
- elements, and other complex genomic architectures, thereby cultivating a comprehensive
- understanding of the entire genomic landscape, including essential 'negative' background sequences.

2.2 Model Architecture Design

- Genos employs a MoE architecture evolved from the Transformer, characterized by 12
- layers, optimized for both performance and efficiency in genomic sequence modeling. The
- 143 Mixture-of-Experts (MoE) architecture is established to provide intrinsic performance benefits
- beyond mere computational efficiency, a principle supporting our genomic model's design.
- Foundational work demonstrated that MoE models achieve superior perplexity and accuracy over

146	dense baselines ur	nder identical	computational b	udgets [10]]. This enhanced	modeling capability	٠,
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- 147 attributed to expert specialization and conditional computation leveraging a vast parameter space, is
- consistently validated across large-scale language models [11] and complex data domains [12].
- 149 Consequently, we directly adopted this well-established architectural design, and the resulting
- 150 performance gains observed in our Genos model align with theoretical expectations and the
- established body of evidence.

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- The model begins with a token embedding layer that converts discrete base tokens into continuous
- vector representations. Following embedding, three root mean square normalization (RMSNorm) [13]
- layers are strategically placed throughout the network to stabilize training by re-scaling inputs to have
- a root mean square of one, without re-centering them around the mean. Between the first and second
- 157 RMSNorm layers, Genos integrates Rotary Position Embedding (RoPE) [8] with an exceptionally
- large base frequency of 50,000,000, enabling it to process ultra-long sequences of up to 1 million
- tokens. Notably, instead of using explicit position embeddings at the input layer, RoPE dynamically
- injects positional information during attention computation by applying rotary transformations to
- query and key vectors. This design offers precise positional awareness while supporting extreme
- 162 context lengths. Complementing RoPE, the model employs a Grouped-Query Attention (GQA) [14]
- mechanism with 16 attention heads sharing 8 key-value groups. This configuration strikes an optimal
- balance between computational efficiency and representational capacity, allowing Genos to process
- long genomic sequences both accurately and efficiently. Genos adopted MoE architecture, which
- 166 consists of a router network and eight expert subnetworks. Each expert subnetwork utilizes SwiGLU
- 167 [15] activation functions, replacing traditional ReLU/GELU for improved expressive capability and
- training stability. The router dynamically selects two out of the eight experts for each token based on
- sequence content, allocating computational resources adaptively (Figure 1). This design enables
- efficient processing of both simple repetitive regions and complex regulatory elements. Finally, a
- linear output layer projects the model's final hidden state into logits over the vocabulary, where the
- softmax function then converts them into a probability distribution for the next token, in accordance
- with the Next Token Prediction (NTP) objective [16]. A key advantage of this model architecture is
- its inherent flexibility, which enables effective adaptation to various downstream applications.

2.3 Pre-training Process and Parameter Optimization

- During the pre-training phase, Genos was trained through the self-supervised paradigm. The model
- employs the NTP objective while producing general genomic representations.
- 178 The model was trained using the Megatron-LM framework [17] across 256 GPUs, employing a
- 179 sophisticated five-dimensional parallelism strategy that combines Tensor Parallelism, Pipeline
- 180 Parallelism, Context Parallelism, Data Parallelism, and Expert Parallelism.
- 181 Training was conducted with a global batch size of 1,024, achieved via gradient accumulation using a
- micro-batch size of 1. The optimization process used the AdamW [18] optimizer with a distributed
- sharded implementation for optimizer states. The learning rate followed a cosine decay schedule,
- starting with a 5% warm-up phase and peaking at 1e-4, accompanied by gradient clipping set at 1.0
- and weight decay of 0.1.
- 186 To address the inherent challenge of expert load imbalance in the MoE architecture—particularly
- pronounced due to the limited vocabulary of genomic sequences (four bases)—we implemented an
- expert load balancing mechanism with auxiliary loss [10] (coefficient 1e-3). This approach prevents

- 189 router collapse and ensures uniform activation of experts across diverse genomic contexts. A Z-loss
- 190 [19] penalty (coefficient 1e-3) applied to router logits to prevent numerical instability and ensure
- smoother training in the MoE components.
- To achieve ultra-long context modeling (up to 1M tokens), we implemented a multi-stage progressive
- training strategy. This approach incorporated three key technical components: training on data with
- 194 progressively increasing context lengths, scheduled learning rate decay to effectively mitigate
- catastrophic forgetting [20], and the application of RoPE-based context window scaling.
- To enhance numerical stability and training quality, mixed-precision training was adopted. This
- involving utilizing BF16 for the majority computations while stricted retaining FP32 precision for
- 198 critical operations, specifically (1) the Softmax function within the attention mechanism, (2) gradient
- accumulation and All-Reduce communications, and (3) MoE routing. Simultaneously, reduced-
- 200 precision matrix multiplication via BF16 was explicitly disabled.
- 201 By integrating GQA and Flash Attention [21], Genos capitalizes on their complementary strengths.
- 202 GQA provides architectural innovations essential for efficient KV caching, while Flash Attention.
- 203 offers an optimized computational kernel for the rapid calculation of attention scores. This synergy
- 204 established a robust foundation for a high-performance large-scale pre-training model capacity for
- 205 extensive context windows.
- 206 Additional optimizations included: Grouped GEMM (General Matrix Multiplication) [22] operations
- for efficient batched expert computation in MoE layers; AllToAll token dispatching [23] for MoE
- 208 communication; Overlapped parameter gathering and gradient reduction to minimize communication
- 209 latency [24]; A cyclic data loader with 8 workers to support continuous data streaming during large-
- scale pretraining.

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2.4 Inference and Downstream Applications

- During inference, Genos leverages its adaptive routing mechanism and GQA to efficiently process
- 213 sequences lengths ranging up to 1 Mb. The model supports three primary modalities: embedding
- 214 generation, sequence generation, and model fine-tuning. For embedding generation, Genos produces
- 215 fixed-dimensional vector representations that capture biological features for tasks such as sequence
- clustering and multi-omics integration. In sequence generation mode, the model functions as an
- 217 autoregressive decoder with sampling strategies including temperature scaling and top-k filtering to
- 218 simulate novel sequences or mutated alleles. On-demand fine-tuning via adapter modules or continual
- 219 learning is available through its hugginface service, enabling customization for specialized tasks such
- 220 as rare disease variant annotation without requiring full model retraining (Figure 1). This aims to
- facilitate deployment across various genomic research and clinical applications.

2.5 Scalable Model Variants: Genos-1.2B and Genos-10B

- 223 To address diverse computational constraints and application scenarios, we developed two versions of
- the Genos model (1.2B and 10B), with architectural details summarized in Table 1. Compared to the
- 225 1.2B variant, the 10B version exhibits substantial improvement across core configuration: the attention
- 226 hidden dimension scales from 1,024 to 4,096, enhancing contextual understanding; the MoE hidden
- 227 dimension per expert quadruples from 4096 to 8192, boosting representational capacity within each
- 228 expert subnetwork; consequently, the total number of parameters rises from 1.25 billion to 10.27 billion,
- expanding model capacity; accordingly, the activated parameter count rises from 0.33 billion to 2.87

billion, reflecting the selective utilization inherent in MoE designs. We trained both versions of the model using the same dataset. For this release, the 10B version has been trained on 2,200B tokens, which is slightly higher than the 1,600B tokens used for the 1.2B version. The contrasting scales of the models define their optimal use cases: the 1.2B version is dedicated to resource-constrained analysis, and most fine-tuning scenarios, while the 10B version is geared towards intensive, high-capacity modeling requirements (e.g., whole-genome structural variation interpretation). A schematic overview of these key capabilities—single-base resolution and ultra-long context modeling—is provided in Figure 2A. The model training process was conducted entirely on the 021 Large Science Model and Zero2X open platform.

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Table 1 Architectural Details of Two Versions of Genos Model

Version	1.2B	10B	
Architecture	M	οΕ	
Number of Total Parameters	1.25B	10.27B	
Number of Activated Parameters	0.33B	2.87B	
Number of Layers	1	2	
Attention Hidden Dimension	1024	4096	
MoE Hidden Dimension (per Expert)	4096	8192	
Number of Attention Heads	1	6	
Number of Experts	8	3	
Selected Experts per Token	2	2	
Vocabulary Size	128(padded)	256(padded)	
Context Length	up to	1M	
Attention Mechanism	GQA&Flash Attention		
Activation Function	SWie	GLU	
Trained Tokens	1600 B	2200 B	

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3. Performance Evaluation

3.1 Benchmark Evaluation and Downstream Application Task

We utilized several standard benchmark datasets to evaluate Genos. We firstly assessed across a 244 suite of established genomics benchmarks, including the Genomics Benchmark (GB), Nucleotide 245

Transformer Benchmark (NTB), and Genomics Long-Range Benchmark (LRB) datasets [25].

From GB, we selected three human-related representative classification tasks: coding versus

noncoding sequence discrimination (demo coding vs intergenomic seqs), enhancer detection (human enhancers cohn), and open chromatin region identification (human ocr ensembl). From

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NTB, we included tasks for splice site recognition (splice sites all) and histone modification

classification (H3, H3K36me3). To evaluate long-range modeling capabilities, four LRB human-

- related tasks were selected, covering enhancer and promoter detection
- 253 (regulatory element enhancer 8K, regulatory element promoter 8K), as well as prediction of
- variant effects on expression (variant effect causal eqtl 8K) and disease pathogenicity
- 255 (variant effect pathogenic clinvar 8K).
- 256 Tasks from GB and NTB involve relatively short DNA sequences (200–600 bp), while the LRB
- 257 framework allows arbitrarily long inputs. We generated 8,192 bp (8K) sequences for all LRB tasks to
- benchmark long-sequence performance. Dataset splits followed the official configurations or, when
- unavailable, chromosome-based partitions. In LRB tasks, chromosome 22 was reserved as a
- validation set. Performance was quantified using the area under the receiver operating characteristic
- 261 curve (AUC) for binary classification tasks and macro-AUC for multi-class settings.
- Next, to further examine scalability to ultra-long inputs, we designed a mutation hotspot classification
- task using data from the Chinese Pangenome Consortium [26]. Sequences of 8,192 bp (8K), 32,768
- bp (32K), and 131,072 bp (128K) were used. Mutation hotspots were identified using a Poisson right-
- tail test, comparing the mutation count of each sequence to the background mean across all segments
- within the same chromosome, with significance determined at FDR < 0.05. The dataset was
- 267 constructed by combining all hotspot sequences with an equal number of randomly selected non-
- 268 hotspot sequences
- 269 Every evaluation task was performed using the sequence model's output embeddings as input to a
- 270 fixed, simple downstream network, enabling direct inter-model comparison.
- 271 In addition to the evaluation tasks, we also conducted two application-level case studies involving
- 272 model fine-tuning tailored to specific application requirements. The primary objective here is not to
- compare intrinsic model capabilities, but rather to illustrate the design of feasible downstream
- applications based on Genos, with the aim of providing case studies for broader practical deployment.
- 275 The Encode and Gtex datasets were employed for tasks such as RNA-seq data generation and gene
- 276 expression analysis. These datasets contain a wealth of single-base transcriptome data from a large
- 277 number of samples, with different cell types and positive and negative strands labeled. By using these
- datasets, we could assess Genos's ability to handle real-world genomic data, learn the underlying
- 279 patterns in gene expression, and generate accurate predictions.
- 280 For evaluating Genos's performance in tasks related to disease association analysis and gene variation
- 281 effect prediction, we adopted datasets related to KEGG and VEP. The KEGG-based dataset contains
- 282 questions related to chromosome information, pathway networks, along with reference and variant
- 283 DNA sequences, and corresponding disease names and reasoning steps. The VEP-based dataset
- 284 focuses on variant effect prediction questions, reference and variant sequences, and the correct
- 285 classification of variant effects. These datasets were carefully constructed to cover a wide range of
- 286 real-world scenarios in genomic research, allowing us to test Genos's performance in complex and
- 287 practical genomic analysis tasks.

3.2 Experimental Results and Comparative Analysis

289 3.2.1 Performance Comparison with Other Models

- We compared Genos with several other relevant models, including GENERator-3b [27], HyenaDNA-
- 291 1M [28], NT-2.5b-multi [29] Evo2-7b, and Evo2 -40b, across different tasks.Both Genos-1.2B and

- Genos-10B demonstrated competitive performance over a wide range of topics and input lengths (Table 2).
- In short-sequence tasks (200–600 bp) (**Table 2**), Genos-10B achieved an AUC of 0.9914 on
- demo_coding_vs_intergenomic_seqs, surpassing models such as GENE-Rator-3B (0.9855),
- 296 HyenaDNA-1M (0.9127), and NT-2.5B-multi (0.9763). On human enhancers cohn, Genos-10B
- 297 reached an AUC of 0.8552, outperforming NT-2.5B-multi (0.7873) and Evo2-7B (0.7733).
- For long-sequence benchmarks (**Table 2**), Genos-10B achieved an AUC of 0.7532 on regulatory
- element_enhancer_8K, comparable to the top-performing models. On variant_effect_
- pathogenic_clinvar_8K, it attained an AUC of 0.9326, markedly exceeding GENE-Rator-3B (0.7206)
- 301 and HyenaDNA-1M (0.6117).
- 302 In the mutation hotspot evaluation (8K–128K inputs) (**Table 2**), Genos-10B consistently achieved the
- 303 highest performance in AUCs. On CPC 131072, it reached 0.9911, outperforming GENE-Rator-3B
- 304 (0.9620) and HyenaDNA-1M (0.9735). Similarly, on CPC_32768, it achieved 0.9625, surpassing
- 305 GENE-Rator-3B (0.9237) and HyenaDNA-1M (0.9064).
- 306 Overall, Genos demonstrated strong and consistent performance across benchmarks of varying
- sequence lengths and biological contexts, highlighting its scalability and robustness from short-range
- 308 genomic classification to ultra-long sequence modeling (Table 2). The overall performance of Genos
- and baseline models across fundamental task categories is summarized visually in Figure 2B and 2C.
- The values shown are averaged from the comprehensive per-task metrics presented in **Table 2**,
- providing a high-level comparison of model capabilities on short- and long-sequence genomic
- 312 understanding.

Table 2 Benchmark Evaluation of Genos Model and Other Genetic Models in Various Tasks^a

	Task	Genos 1.2B	Genos 10B	GENERat or-3b	HyenaD NA-1M	^b NT- 2.5b- multi	°Evo2- 7b	°Evo2- 40b
Short	demo_coding_vs_intergenomi c_seqs	0.9708	0.9914	0.9855	0.9127	0.9763	0.9824	0.9886
sequence evaluation	human_enhancers_cohn	0.8715	0.8552	0.8181	0.7799	0.7873	0.7733	0.7756
(sequence	human_ocr_ensembl	0.7569	0.7623	0.7270	0.6916	0.7285	0.7505	0.7635
length	splice_sites_all	0.7819	0.7990	0.8071	0.7110	0.8603	0.8747	0.9138
200- 600bp)	H3	0.8944	0.9400	0.9163	0.8722	0.9371	0.9140	0.9311
оооор)	H3K36me3	0.6883	0.7658	0.8247	0.6787	0.8288	0.8615	0.8823
Mutation	CPC_131072	0.9872	0.9911	0.9620	0.9735	/	/	/
hot spot evaluation	CPC_32768	0.9440	0.9625	0.9237	0.9064	/	0.9504	0.9611
(sequence length: 8K ~ 128K)	CPC_8192	0.9093	0.9522	0.9315	0.8914	/	0.9425	0.9401
Long	regulatory_element_enhancer _8K	0.7469	0.7532	0.7390	0.7282	/	0.7454	0.7527
sequence evaluation	$\begin{array}{ccc} regulatory_element_promoter \\ -8K \end{array}$	0.9221	0.9249	0.9195	0.8890	/	0.9255	0.9227
(sequence length:	variant_effect_causal_eqtl_8 K	0.6990	0.6773	0.6920	0.6887	/	0.7039	0.7054
8K)	variant_effect_pathogenic_cli nvar_8K	0.6907	0.9326	0.7206	0.6117	/	0.7308	0.9167

- a. Public models of HyenaDNA, Nucleotide Transformer (NT), and other versions in the GENERator series
- have also been tested. Due to space limitations, the evaluated models not listed include GENERator-1.2b,
- 316 HyenaDNA-32k, HyenaDNA-450k, NT-500M, and Evo2-1b. Here, only the best-performing models from each
- 317 series are shown.

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- b. For NT public models, the maximum acceptable input length is 6000, making them unavailable for tasks with
- 319 input lengths of 8K or more.
- 320 c. Evo2 7b and 40b models cannot perform inference for 128K or longer sequences under the HuggingFace
- framework due to resource constraints.

4. Case Studies

4.1 RNA-seq Profiles Prediction Case

4.1.1 Data Preparation and Preprocessing Steps

- 325 In this case, we fine-tune Genos after modifying its output head with a task-specific
- architecture to predict single-base resolution RNA-seq profiles from DNA sequences across
- diverse cell types and tissues. In the same way as AlphaGenome, the training data were
- sourced from ENCODE [30] and GTEx [31], yielding a total of 667 metadata groups of
- 329 single-base transcriptome samples. The data was preprocessed by first normalizing all
- BigWig files to a common scaling factor and then averaging the expression values across
- samples within each group to generate an average normalized RNA-seq profile for every
- distinct biological context. The model was trained on paired data, using hg38 reference
- genome sequences as input and the corresponding averaged RNA-seq profiles as output
- targets. Considering fine-tuning costs and the consistency of local sequence predictions, we
- set the sequence window length to 32 kb, with a 16 kb overlap between adjacent windows.
- Data sampling spanned all positions across chromosomes 1–22.
- This data preparation and preprocessing strategy aims to provide high-quality and consistent
- data for subsequent model training, ensuring that the model can effectively learn the
- underlying relationships between genomic sequences and their corresponding transcriptomic
- 340 expressions.

4.1.2 Network Architecture and Training Process

- Full fine-tuning is conducted on the Genos-1.2B model for each RNA-seq profile. The
- downstream task head employs a convolutional architecture comprising three 1D
- convolutional layers, configured with (kernel size, padding, dilation) pairs of (3, 1, 1), (3, 2,
- 2), and (1, 0, 1), respectively. Channel dimensions are progressively reduced from 1024 to
- 256, 256 to 64, and 64 to 1. Each convolutional layer is followed by batch normalization,
- GELU activation, and dropout regularization (dropout rate = 0.1). The final output is scaled
- via a learnable weight parameter and transformed through a Softplus activation function to
- 349 enforce non-negative predictions.
- We employed MSE as the loss function for this token-level regression task. To ensure
- training stability, we implemented a data scaling strategy similar to AlphaGenome: a square-
- 352 root-based smooth clipping and power transformation were applied to compress signal values
- during training, and the inverse operations were performed when inference.
- For optimization, we employed the Adafactor optimizer with a cosine annealing learning rate
- 355 scheduler and a linear warmup phase covering 5% of the total training steps. The global batch
- size was set to 256, and each model was trained for 60 epochs totally.

4.1.3 Evaluation and Result Analysis

- To assess the fidelity of RNA-seq profile predicted by fine-tuned Genos, we quantified the
- consistency between model-generated and experimentally derived RNA-seq profiles across
- two cell types: the human B lymphoblastoid cell line (GM12878, EFO:0002784) and natural
- 361 killer cell (CL:0000623). For each cell type (stratified by DNA strand orientation, "+" or
- 362 "-"), we calculated log1p-transformed Pearson correlation coefficients across three genomic
- scopes: whole genome, gene region, and gene expression matrix.
- As summarized in Table 3, Genos demonstrated strong agreement with experimental RNA-
- seq results across all scenarios. In GM12878 cells, log1p Pearson correlations reached 0.9335
- (whole genome, + strand), 0.9334 (gene region, + strand), and 0.8641 (gene expression, +
- strand); for the strand, these values were 0.9182 (whole genome), 0.9274 (gene region), and
- 368 0.9081 (gene expression). In natural killer cells, the model achieved correlations of 0.9084
- (whole genome, + strand), 0.9036 (gene region, + strand), 0.9267 (gene expression, + strand),
- and 0.8562 (whole genome, strand), 0.8542 (gene region, strand), 0.8969 (gene
- 371 expression, strand).

- These high correlation values are further corroborated by visual inspection of RNA-seq signal
- tracks (Figure 3). The figure illustrates a 32 kb genomic region (chr19:39,407,000
- 374 39,439,000) with annotated genes (e.g., ZPBP, RPL36C2, MPL) at the top. Different colored
- tracks represent Genos-generated total RNA-seq signals for EFO:0002784 (human B
- lymphoblastoid cell line GM12878) (blue tracks) and CL:0000623 (natural killer cell)
- (orange/green/red tracks) across positive (+) and negative (-) strands. For GM12878
- 378 (positive strand), signal peaks align precisely with the exonic regions of RPL36C2, reflecting
- 379 strand-specific transcriptional activity. In natural killer cells, signals concentrate near the
- MPL locus, and their strand orientation matches the known transcriptional direction of MPL
- transcripts. These visual patterns confirm that Genos not only achieves high quantitative
- 382 correlation (Table 3) but also recapitulates cell type–specific and strand-specific
- transcriptomic landscapes, with signal distributions that accurately mirror gene structure and
- 384 cell type–dependent expression patterns.
- Overall, both quantitative and visual evidence validate Genos as a reliable tool for in silico
- RNA-seq data generation, capturing both global transcriptional patterns (evidenced by whole-
- genome and gene-region correlations) and gene-specific expression dynamics (reflected in
- gene expression matrix correlations and signal track alignments). While slight reductions in
- correlation within gene expression analyses may reflect residual challenges in modeling fine-
- 390 grained transcriptomic variation, the strong performance across modalities supports the
- 391 model's utility for transcriptomic research.
- 392 It is noteworthy that our preliminary fine-tuning of the larger Genos-10B parameter model on
- this task, though currently limited to chromosome 19, already indicates a performance superior
- 394 to the specialized model AlphaGenome (Supplementary Figure S1 and Table S1). As the
- 395 genome-wide fine-tuning for the 10B model is ongoing and not yet ready for full release, we
- conservatively report the results of the fully evaluated Genos-1.2B model in the main text.

Table 3 Consistency between RNA-seq data of two cell types generated by the Genos-1.2B model and the actual results

Туре	Cell Types	Genes chain	log1p Pearson (Whole genome)	log1p Pearson (Gene region)	log1p Pearson (Gene expression)
total RNA-seq	GM12878 (EFO:0002784)	+	0.933467	0.933387	0.8641
total RNA-seq	GM12878 (EFO:0002784)	-	0.918187	0.927362	0.9081
total RNA-seq	natural killer cell (CL:0000623)	+	0.908418	0.903551	0.9267
total RNA-seq	natural killer cell (CL:0000623)	-	0.856171	0.854174	0.8969

4.2 Text-genome Model Fusion Case

4.2.1 Project Overview and Data

To evaluate the performance of a multimodal large language model (combining a genome model and a text model) for predicting genetic diseases caused by gene variants, we follow the architecture [32], which is capable of processing raw DNA sequences while leveraging the reasoning capabilities of large language models to generate biologically consistent explanations and predictions.

The data used to show the text-genome model fusion is derived from the KEGG task introduced in the Bioreason paper [32] This task integrates KEGG pathway information with clinical mutation data through a multi-stage pipeline, employing a standardized symbolic system to represent various molecular interactions and providing reference sequences for comparison with mutated sequences (**Figure 4**). The KEGG dataset comprises 1,449 entries spanning 37 distinct diseases, and is partitioned into training, validation, and test sets in an 8:1:1 ratio. Each input consists of a problem description, reference gene sequences, and corresponding mutated gene sequences. The outputs include both reasoning steps and disease classification predictions.

4.2.2 Data Preprocessing and Model Training

The data processing workflow adheres to the KEGG processing steps described in Bioreason [32]. The maximum DNA sequence length is limited to 1,024 base pairs. The goal of this model training is to achieve efficient alignment between DNA sequences and natural language. We utilized two series of text models. The first is the Qwen3 [33] series, which includes the Qwen3-1B, Qwen3-4B, and Qwen3-8B models. The second is the 021 Science Foundation Model, which is a large language model trained on extensive scientific corpora with profound scientific cognition. The model training used the AdamW optimizer with a learning rate of 5×10^{-5} and a weight decay of 1×10^{-2} . Gradient accumulation was set to 8 steps, and a random seed of 23 was used for reproducibility. LoRA adapters were applied with a rank of 32, an alpha value of 64, and a dropout rate of 0.05. Fine-tuning was performed on the text model (text_model_finetune: True), while the DNA model was frozen. These

settings were designed to optimize the training process, balancing model accuracy and computational 429 430 efficiency through proper regularization and parameter tuning.

4.2.3 Evaluation Indicator Scheme and Results

To evaluate the multi-label classification task, we employed standard metrics, including Accuracy, Macro Precision, Macro Recall, and Macro F1-score. These metrics were selected to assess multi-label disease prediction performance, while accounting for potential class imbalance. The results indicate that model performance varied across different architectures and input combinations. For instance, among genome-only models, the Genos-10B model achieved an accuracy of 92.07%, a Macro F1-score of 72.59%, a Macro Precision of 75.46%, and a Macro Recall of 74.15%. As shown in Table 4, in the genome-text models, the Genos-1.2B + 021-8B model achieved an accuracy of 98.28% and a Macro F1-score of 90.37%, demonstrating its effectiveness in processing raw DNA sequences and accurately predicting disease outcomes.

Table 4 Evaluation Indicators of the Genos Model + Text Model Diagnostic Model

Model_type	Model	Accuracy	F1- score	Precision	Recall
	Genos-10B	92.07%	72.59%	75.46%	74.15%
	Genos-1.2B	91.72%	72.89%	75.24%	72.95%
Genome	Evo2-1.2B	88.28%	72.43%	75.23%	69.83%
	NT-2.5b-multi	86.55%	69.76%	73.23%	66.62%
	HyenaDNA-1M	50.00%	11.11%	9.22%	14.98%
	Genos-1.2B + 021-8B	98.28%	90.37%	97.87%	90.15%
	Evo2-1.2B + 021-8B	97.59%	90.96%	98.49%	90.82%
	HyenaDNA-1m + Qwen3-8B	97.58%	95.61%	100%	94.79%
	Evo2-1.2B + Qwen3-4B	97.24%	86.30%	86.75%	87.25%
	Genos- $10B + 021-8B$	97.23%	92.32%	100.00%	90.51%
	Genos-1.2B + Qwen3-4B	96.90%	93.24%	100.00%	91.15%
	NT-2.5b-multi + Qwen4B	96.90%	89.03%	90.99%	89.38%
	HyenaDNA-1M + 021-8B	96.55%	93.34%	97.03%	92.86%
Genome-text	Evo2-1.2B + Qwen3-8B	96.21%	93.53%	100.00%	91.47%
	Genos-10B + Qwen3-4B	96.21%	91.75%	100.00%	90.30%
	HyenaDNA-1M + Qwen3-4B	96.21%	89.55%	99.94%	87.02%
	HyenaDNA-1M + Qwen3-1B	93.45%	93.12%	99.05%	91.29%
	Genos-1.2B + Qwen3-1B	91.38%	83.08%	99.57%	79.50%
	Evo2-1.2B + Qwen3-1B	90.42%	75.62%	77.42%	73.91%
	Genos-10B + Qwen1B	88.97%	79.24%	98.22%	76.99%
	NT-2.5b-multi + Qwen1B	88.42%	72.13%	75.42%	71.91%

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5. Deployment and Application Prospects

5.1 Current Deployment Status and Usage

- 449 Currently, Genos is in the R&D and optimization phase. It is mainly supporting internal scientific
- research, providing a powerful tool for researchers within the organization to conduct in-depth
- 451 genomic studies. Genos is designed to be highly adaptable to mainstream GPU environments, with no
- 452 special hardware restrictions. This compatibility ensures that it can be easily integrated into existing
- research setups, reducing the barriers to its utilization.
- 454 Adhering to the concept of open science, Genos has deployed cloud reasoning services on the DCS-
- Cloud platform, thereby constructing a "cloud lab" for genomic intelligence analysis. This open-
- ecology initiative has far-reaching implications.

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- Researchers can upload their data through an intuitive interface. Once the data is uploaded, Genos can
- 458 perform a full-process analysis, starting from mutation function annotation. Mutation function
- 459 annotation helps in understanding the biological significance of genetic mutations, whether they are
- 460 benign, pathogenic, or have some other functional implications. The analysis then extends to
- 461 phenotype prediction, which is a crucial step in connecting genetic information to observable traits.
- This decentralized computing power support model breaks the shackles of local computing power and
- 463 algorithm deployment limitations. Researchers from all over the world can now share the predictive
- power of this leading -edge model. For example, a research team in a resource-limited region can
- access Genos through the cloud service, enabling them to conduct high-level genomic analysis that
- 466 was previously out of reach due to lack of local computational resources. This accelerates the
- transition from genomic discovery to clinical applications, as more research can be carried out and
- validated, bringing genomic insights closer to patient care.

5.2 Future Application Potential in Biomedicine

- 470 In the field of precision medicine, Genos holds great promise. It can analyze an individual's genomic
- data to identify disease-related genetic markers with high precision. For example, in cancer diagnosis,
- 472 Genos can analyze tumor-associated genomic variations, predict the aggressiveness of the cancer, and
- suggest personalized treatment plans. By accurately predicting the response of different patients to
- 474 various drugs based on their genetic makeup, Genos can help doctors select the most effective
- treatment options, minimizing the risk of adverse reactions and improving treatment outcomes.
- For group health monitoring, Genos can analyze the genomic data of a large population. It can
- 477 identify genetic factors associated with common diseases in the population, such as cardiovascular
- diseases, diabetes, and neurodegenerative disorders. This information can be used to develop
- 479 preventive strategies, such as targeted health education, lifestyle interventions, and early-detection
- 480 screening programs for high-risk individuals.
- 481 In developmental biology, Genos can contribute to understanding the genetic basis of embryo
- development. By analyzing the genomic changes during different stages of embryo development, it
- 483 can uncover the regulatory mechanisms that control cell differentiation, organ formation, and overall
- 484 development. This knowledge can help in diagnosing and treating developmental disorders and also
- 485 provide insights into reproductive medicine, such as improving in vitro fertilization techniques. As
- 486 Genos continues to optimize and iterate, its application potential in these biomedical fields will

continue to expand, laying a solid foundation for the development of a more comprehensive and

488 effective healthcare system.

6. Conclusion

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6.1 Summary of Research Findings

- 491 Genos represents a significant advancement in genomic intelligence analysis. Specifically, its Mixture-of-
- 492 Experts (MoE) architecture effectively addresses the computational challenges inherent in ultra-long
- 493 sequence modeling at single-nucleotide resolution. By introducing strategies such as ultra-long sequence
- 494 parameterization, multi-dimensional parallel computing, and complementary attention mechanisms, Genos
- 495 successfully overcomes the limitations of traditional models in handling million-base sequences. The expert
- 496 load balancing mechanism, mixed-precision training strategy, and dynamic routing architecture further
- 497 enhance the model's training stability and inference efficiency.
- 498 In terms of performance, Genos outperforms existing models in various benchmark tasks. In addition
- 499 to the existing benchmark comparisons, we designed two specific tasks focused on ultra-long
- sequence modeling. Across these tasks, the Genos model exhibited a clear positive correlation
- between sequence length and prediction accuracy. In contrast, other models were either unable to
- 502 process sequences of such lengths or did not demonstrate this property of performance scaling with
- 503 increased sequence context. This finding thus provides empirical evidence for the necessity of longer
- 504 context windows.
- 505 The application cases of Genos further validate its utility. In RNA-seq data generation, Genos can
- accurately predict gene expression levels, as demonstrated by high Pearson correlation coefficients
- 507 between predicted and true expression values. In the omics + text interactive disease diagnosis
- 508 project, Genos, when combined with a large-scale language model, achieves high accuracy in gene
- variation effect prediction and disease association analysis, with accuracy rates reaching up to 99.31%
- 510 in some cases.

511 **6.2 Limitations and Future Work**

- 512 The Genos model has several limitations that must be addressed in future work. Firstly, computational
- 513 efficiency requires optimization. Although the architecture is designed for effective resource
- allocation, there is still potential to significantly reduce the computational cost during training and
- 515 inference, particularly when handling massive datasets. Secondly, the capability for cross-modal data
- fusion needs enhancement. While Genos shows initial promise with genomic data, deeper integration
- of multi-omics data, such as proteomics and metabolomics, alongside phenotypic information, is
- 518 essential to achieve a more comprehensive understanding of complex biological processes and gene-
- 519 environment interactions.

- 521 Future model development will involve continuous training with an increasingly diverse set of
- 522 genomic data, with the primary objective remaining a deeper comprehension of the human genome
- 523 and superior performance in corresponding analytical applications. Furthermore, the integration of
- 524 other multi-omics datasets with the Genos genomic model is anticipated to offer substantial benefits
- for downstream research and practical applications. Additionally, while Genos's architectural features
- (e.g., long-context attention and MoE) are designed to facilitate contextual learning, a comprehensive

527	benchmark evaluating its performance across a wide array of human tissues—such as predicting RNA
528	expression or chromatin accessibility in diverse GTEx or ENCODE contexts—remains an important
529	area for future validation and will be a focus of subsequent studies.
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531	6.3 Significance of Genos for Genomics Development
532	Genos is expected to have a substantial impact on the trajectory of genomics research. It marks a
533	paradigm shift from traditional data-driven genomics research to an foundation model-based
534	approach. By providing a powerful tool for accurate and efficient genomic analysis, Genos enables
535	researchers to gain deeper insights into the genetic basis of diseases.
536	Within the domain of precision medicine, Genos may play a crucial role in disease risk prediction,
537	personalized diagnosis, and treatmen stratificationt. Its capacity to analyze genomic data at a high
538	level of accuracy can aid in identifying disease-associated genetic variants, predicting the efficacy of
539	drugs, and developing personalized treatment plans. This may lead to more effective and targeted
540	medical interventions, reducing the cost and side-effects associated with traditional treatment
541	methods.
542	Moreover, Genos contributes to a more comprehensive understanding of life processes. By decoding
543	the intricate genomic information, it paves the way for advancements in fields such as developmental
544	biology, evolutionary biology, and synthetic biology. Overall, Genos is a key step towards realizing
545	the full potential of genomics in improving human health and understanding the mysteries of life.
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547	Availability of Source Code and Requirements
548	Project name: Genos
549 550	Project homepage: https://github.com/BGI-HangzhouAI/Genos & https://huggingface.co/BGI-HangzhouAI
551	Operating system(s): Platform independent
552	Programming language: Python
553	Other requirements: pytorch 7.1 or higher, transformers 4.52.4 or higher
554	License: MIT license
555	
556	Data Availability
557 558	To facilitate reproducible research and community collaboration, all resources for the Genos model are publicly accessible. Pre-trained model weights, inference code, and detailed documentation are
559	released on GitHub (https://github.com/BGI-HangzhouAI/Genos) and the Hugging Face Hub
560	(https://huggingface.co/BGI-HangzhouAI). These resources enable researchers to fine-tune Genos for
561	specialized genomic tasks or integrate it into custom bioinformatic workflows. The model is
562	distributed under the MIT License, permitting unrestricted use, modification, and redistribution for
563	both academic and commercial purposes. For users seeking scalable cloud-based inference, Genos is also deployed on the BGI DCS Cloud platform, with dedicated APIs to support end-to-end genomic
564 565	analysis without local computing infrastructure.
566	amaysis without local companing infrastructure.
567	Disclosure of use of AI-assisted tools including generative AI
568	In the preparation of this manuscript, an AI-assisted tool (Doubao) was utilized to support the
569	optimization of academic writing structure (e.g., organizing the logical flow of the Methodology
570	section and Abstract), and refine the expression of technical content. All content generated or
571	optimized with the assistance of this tool underwent a thorough process of review, verification, and
572 573	revision by the authors to ensure accuracy, academic rigor, and consistency with the study's original
574	findings.
575	Acknowledgement
576	We would like to acknowledge the Human Genome Structural Variation Consortium (HGSVC) and
577	the Human Pangenome Reference Consortium (BioProject ID: PRJNA698480), as well as their
578	funder, the National Human Genome Research Institute (NHGRI). We thank Zhejiang Lab for
579	developing the 021 Science Foundation Model, which is scheduled to be released at a later date.
580	The model training process was conducted entirely on the 021 Large Science Model and Zero2X open
581	platform.
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References

- 586 1. Brixi, G., et al., *Genome modeling and design across all domains of life with Evo 2.* bioRxiv, 2025: p. 2025.02.18.638918.
- Avsec, Ž., et al., *AlphaGenome: advancing regulatory variant effect prediction with a unified DNA sequence model.* bioRxiv, 2025: p. 2025.06.25.661532.
- Hickey, G., et al., *Pangenome graph construction from genome alignments with Minigraph-Cactus*. Nat Biotechnol, 2024. **42**(4): p. 663-673.
- 592 4. Vollger, M.R., et al., *Increased mutation and gene conversion within human* segmental duplications. Nature, 2023. **617**(7960): p. 325-334.
- 594 5. Liao, W.W., et al., *A draft human pangenome reference*. Nature, 2023. **617**(7960): p. 312-324.
- 596 6. Fairley, S., et al., *The International Genome Sample Resource (IGSR) collection of*597 open human genomic variation resources. Nucleic Acids Res, 2020. **48**(D1): p. D941598 D947.
- 599 7. Jacobs, R.A., et al., *Adaptive Mixtures of Local Experts.* Neural Computation, 1991; 600 p. 79-87.
- 8. Su, J., et al., *RoFormer: Enhanced transformer with Rotary Position Embedding.*Neurocomputing, 2024. **568**: p. 127063.
- 9. Vaswani, A., et al., *Attention is all you need.* Advances in neural information processing systems, 2017. **30**.
- 505 10. Shazeer, N.M., et al., *Outrageously Large Neural Networks: The Sparsely-Gated Mixture-of-Experts Layer.* International Conference on Learning Representations, 2017. **1701.06538**.
- Fedus, W., B. Zoph, and N. Shazeer, Switch Transformers: Scaling to Trillion
 Parameter Models with Simple and Efficient Sparsity. Journal of Machine Learning
 Research, 2022. 23.
- Shi, X.L., et al., *Time-MoE: Billion-Scale Time Series Foundation Models with Mixture of Experts.* International Conference on Learning Representations, 2024. **2409.16040**.
- Thang, B. and R. Sennrich, Root Mean Square Layer Normalization. arXiv e-prints,
 2019: p. arXiv:1910.07467.
- Ainslie, J., et al., GQA: Training Generalized Multi-Query Transformer Models from
 Multi-He ad Checkpoints. The 2023 Conference on Empirical Methods in Natural
 Language Processing, 2023.
- 519 15. Zhai, X., et al., *Scaling Vision Transformers*. Proceedings of the IEEE/CVF Conference on Computer Vision and Pattern Recognition (CVPR): p. 12104-12113.
- 621 16. Radford, A. and K. Narasimhan, *Improving Language Understanding by Generative Pre-Training.* 2018.
- Shoeybi, M., et al., *Megatron-lm: Training multi-billion parameter language models using model parallelism.* arXiv preprint arXiv:1909.08053, 2019.
- Loshchilov, I. and F. Hutter, *Decoupled weight decay regularization.* arXiv preprint arXiv:1711.05101, 2017.
- 527 19. Zoph, B., et al., *ST-MoE: Designing Stable and Transferable Sparse Expert Models.* arXiv preprint arXiv:2202.08906, 2022.
- Wang, X, et al., Learning Dynamics in Continual Pre-Training for Large Language Models. Forty-second International Conference on Machine Learning, 2025.
- Dao, T., et al., FlashAttention: Fast and Memory-Efficient Exact Attention with IO-Awar eness. Advances in Neural Information Processing Systems 35 (NeurIPS 2022) 2022.
- Zhai, Y., et al. ByteTransformer: A High-Performance Transformer Boosted for
 Variable-Length Inputs. in 2023 IEEE International Parallel and Distributed
 Processing Symposium (IPDPS). 2023.

- Hwang, C., et al., *Tutel: Adaptive Mixture-of-Experts at Scale.* Proceedings of Machine Learning and Systems, 2022. **5, 269-287**.
- 639 24. Liu, J., et al., A Survey on Inference Optimization Techniques for Mixture of Experts M odels. arXiv preprint arXiv:2412.14219, 2024.
- Trop, E., et al., *The Genomics Long-Range Benchmark: Advancing DNA Language Models*. 2025.
- 643 26. Gao, Y., et al., *A pangenome reference of 36 Chinese populations.* Nature, 2023. **619**(7968): p. 112-121.
- Wu, W., et al., *GENERator: A Long-Context Generative Genomic Foundation Model.* arXiv preprint arXiv:2502.07272, 2025.
- Nguyen, E., et al., *Hyenadna: Long-range genomic sequence modeling at single nucleotide resolution.* Advances in neural information processing systems, 2023. **36**: p. 43177-43201.
- Dalla-Torre, H., et al., *Nucleotide Transformer: building and evaluating robust* foundation models for human genomics. Nat Methods, 2025. **22**(2): p. 287-297.
- 652 30. Consortium, E.P., *An integrated encyclopedia of DNA elements in the human genome.* Nature, 2012. **489**(7414): p. 57-74.
- Kim-Hellmuth, S., et al., *Cell type-specific genetic regulation of gene expression across human tissues*. Science, 2020. **369**(6509).
- 656 32. Fallahpour, A., et al., *BioReason: Incentivizing Multimodal Biological Reasoning within a DNA- LLM Model.* arXiv preprint arXiv:2505.23579, 2025.
- 33. Yang, A., et al., Qwen3 technical report. arXiv preprint arXiv:2505.09388, 2025.

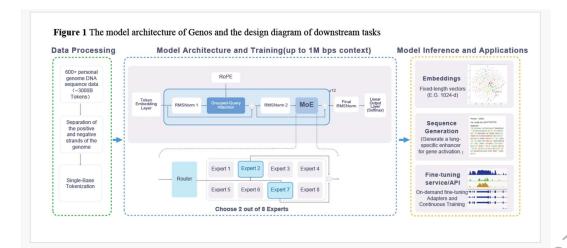


Figure 2. Architectural overview and benchmark performance of the Genos model.

- (A) Schematic illustration of Genos's core capabilities: 1 Mb ultra-long context window and single-base precision, enabling the model to analyze genomic sequences from the nucleosomal level down to individual nucleotides for capturing long-range functional regulation.
- (B) Short-sequence task performance: precise genomic element annotation. Bar plots show the accuracy of Genos and baseline models on tasks including enhancer, exon, intron, and mutation hotspot recognition. Results are averaged across the respective task categories from the comprehensive benchmark in Table 2.
- (C) Long-sequence task performance: capturing long-range regulatory signals. Bar plots compare the accuracy of models on predictions requiring the understanding of long-range interactions.

